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(54) Title: PHARMACEUTICAL COMPOSITION, CONTAINING FRAGMENTS OF AN ANTIGENIC PROTEIN ENCODING DNA ENDOWED WITH ANTI-TUMOR EFFECT		
<p>(57) Abstract</p> <p>Provided herein is a pharmaceutical composition containing one or more DNA molecules encoding fragments of a protein overexpressed in tumor cells, in order to induce an anti-tumor Ag-specific immune response, in association with suitable excipients and adjuvants.</p>		

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PHARMACEUTICAL COMPOSITION, CONTAINING FRAGMENTS OF AN ANTIGENIC PROTEIN ENCODING DNA ENDOWED WITH ANTI-TUMOR EFFECT.

Field of the invention

5 The invention relates to a pool of DNA plasmid constructs containing the sequences of human MUC-1 encoding fragments and to a pool of DNA plasmids in which the fragments themselves are preceded by the sequence encoding a protein consisting of human ubiquitin fused to a bacterial LacI fragment. The invention
10 further relates to their use in the preparation of pharmaceutical compositions for use as DNA anti-tumor vaccines.

Background art

The invention provides an anti-tumor therapy based on the induction or activation of the immune response able to bring
15 about tumor rejection. The validity of such an idea is demonstrated from the first clinical results; for example, patients treated with a viral vaccine containing the Carcinoembryonic Antigen (CEA) encoding sequences demonstrated immune system activation against this antigen (Tsang KY et al.
20 J. Natl. Cancer. Inst. 87: 982, 1995).

The activation of an immune anti-tumor response is achievable through four different approaches:

- a) *Ex vivo* engineering of patient tumor cells in order to make them more immunogenic and suitable as a vaccine;
- 25 b) *Ex vivo* engineering of patient immune cells in order to pre-activate an *in vitro* immune response.
- c) Inoculation of naked or liposome capsulated or viral particle integrated (retrovirus, vaccinia virus, adenovirus, etc.) DNA encoding tumor associated antigens;
- 30 d) Treatment with recombinant or synthetic soluble tumor antigens conjugated or mixed with adjuvants.

The first two approaches consist of the engineering of every single patient cell and are limited in that they are necessarily patient-specific, while the latter two are aimed to

obtain products comparable to a traditional drug.

The new vaccination methods reflect the development of new technologies. The recent indications coming from the experimentation on DNA naked vaccines that induce either a persistent antibody or a cell immune response, make the traditional protein subunit vaccines constituted of certain specific peptides, inducing a lymphocyte population, obsolete. Intramuscularly or intradermically injected proteins, encoded by naked DNA, induce a cytotoxic-specific response as well as a helper response. This powerful combination is extremely effective but the underlying mechanism is not completely clarified yet. Muscle cells express class I MHC antigens at low levels only, and do not apparently express class II antigens or co-stimulatory molecules. Consequently, transfected muscle cells are unlikely to play an important role in the onset of the immune response per se. Recent data show that Antigen Presenting Cells (APC), such as macrophages or dendritic cells, play a fundamental role in capturing the myocyte released antigen and in the subsequent processing and presenting of the respective peptides in the context of the class I and II molecules, thus inducing a CD8+ cell activation with cytotoxic activity as well as activation of the CD4+ cells co-operating with B lymphocytes in eliciting the antibody response (Corr M et al *J. Exp. Med.* 184:1555, 1996) (Tighe, H. et al. *Immunology Today* 19:89, 1998).

Furthermore, the use of cytokines is known to improve the therapeutic effect deriving from immunization with DNA. Cytokines can be administered in the form of exogenous proteins as reported in Irvine et al., *J. Immunol.* 156: 238, 1996. An alternative approach is represented by the contemporaneous inoculation of both the tumor antigen or the desired cytokine encoding plasmids, thus allowing the cytokine to be produced in situ (Kim JJ et al. *Immunol* 158: 816, 1997).

The active immunization approach of the present invention is based on the use of DNA vectors as vaccines against the MUC-1

human antigen or Polymorphic Epithelial Mucin (PEM), overexpressed in tumor cells. MUC-1 is an epithelial luminal surface glycoprotein (Patton S. et al. *BBA* 1241:407, 1995). In the cell transformation process this glycoprotein loses the apical localization and its expression level rises dramatically. The protein function consists of protecting the luminal surfaces, for example in the mammal gland, ovary, endometrium, colon, stomach, pancreas, bladder, kidney, etc. A glycosylation defect is reported that makes tumor cell associated MUC-1 antigenically different from normal cell associated MUC-1. This phenomenon causes tumor MUC-1 to expose the antigen epitopes that are normally masked by the sugar moieties in the normal cell expressed MUC-1. This characteristic makes tumor MUC-1 particularly interesting in an induction of a tumor specific antibody response (Apostolopoulos V. et al. *Crit. Rev. Immunol.* 14:293, 1994).

As an objective, the vaccination is aimed at inducing immune responses against tumor cells expressing MUC1 at high levels, preserving at the same time the low expressing normal epithelia. The DNA vaccination relies upon the entrance of a gene or portions thereof inside the body cells followed by transcription and translation of the inserted sequence and thus the intracellular synthesis of the corresponding polypeptide. An important advantage of this system is that the neo-synthesized protein is naturally processed inside the cell and the produced peptides are associated with the Major Histocompatibility Complex class I molecules (MHC-I). The MHC/peptide complexes are therefore naturally exported to the cell surface where they can be recognized by the immune system CD8+ cytotoxic cells. Only the polypeptides synthesized inside the cell are then processed and presented in association with the MHC class I molecules, thus making it the only mechanism to stimulate, a specific cytotoxic response. Vaccination systems based on protein or peptide administration are usually more effective in stimulating

the antibody immune response which, to date, has been shown to be ineffective in rejecting tumor cells. Current gene therapy techniques rely upon DNA packaging in recombinant viral vectors (retrovirus and adenovirus). The naked DNA administration is much more advantageous in terms of effectiveness and safety compared to viral vector therapies (Kumar V and Sercarz E. *Nature Med.* 2: 857, 1996; McDonnell WM et al., *New England J. of Med.* 334: 42, 1996). In fact naked DNA is unable either to duplicate or integrate in the host tissue DNA and does not induce the immune response to viral proteins.

The use of the ubiquitin to enhance the neo-synthesized protein processing and thus cytotoxic lymphocyte induction was recently reported (Rodriguez F. et al., *J. Virology* 71: 8497, 1997). The use of ubiquitin in order to generate proteins with an N-terminal amino acid, making them unstable and thus prone to enhanced degradation, had been previously reported (Bechmair A. et al., *SCIENCE* 234: 179, 1986). The higher instability of these proteins was subsequently related to enhanced intracellular processing and presentation of model proteins by MHC-1 (Grant E P et al., *J. Immunol.* 155: 3750, 1995) (Wu Y and Kipps T.J., *J. Immunol.* 159: 6037, 1997).

The use of single constructs containing partial antigen encoding DNA fragments (influenza virus nucleoprotein), having a higher antigenic presentation efficiency compared to the analogues with the whole antigenic sequence, in DNA vaccination was reported (Anton L. C. et al., *J. Immunol.* 158: 2535, 1997). Furthermore the processing of intracellular proteins and presentation of the respective peptides by MHC class I proteins in physiologic conditions, underlie the mechanism of immunological surveillance. For a given protein and a specific MHC context, there are peptide fragments termed dominants (i. e. prevailing on subdominants or cryptics), which are unable to generate any immune response because they are recognized as "self". It has now been outlined, according to an aspect of the

present invention, that an approach aimed at supporting the non-dominant epitope presentation by the administration of a mix of antigen protein fragments is able to elicit a surprising cytotoxic immune response.

5 Description of the invention

It has now been found that DNA molecules, encoding fragments of a protein overexpressed in tumor cells, can be conveniently used to induce an antigen-specific anti-tumor immune response.

10 The invention relates particularly to a pharmaceutical composition containing one or more DNA encoding Mucin (MUC-1) protein fragments.

 The DNA used in the present invention can be plasmid or viral DNA, preferably plasmid DNA obtained employing the pMRS30
15 expression vector described in fig. 13.

 The compositions according to the invention contain preferably at least two DNA fragments of the Mucin (MUC-1) or of another protein overexpressed in tumor cells.

 The compositions according to the invention contain
20 preferably at least four fragments, each ranging from 200 to about 700 nucleotides, each sequence being juxtaposed and possibly partially overlapping, from about 50 to about 150 nucleotides, at the 3' and/or 5' end of the adjacent one.

 The DNA fragments according to the invention can be
25 possibly preceded at the 5' end by a ubiquitin encoding DNA sequence and possibly also by a LacI portion of Escherichia coli.

 The invention relates also to new DNA fragments and to the use of Mucin-1 fragments defined above in the medicine and anti-
30 tumor vaccine preparation.

Description of the figures

 Fig. 1

 Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS166 expression

vector. This DNA includes the sequence corresponding to nucleotides 136-339 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by the two translation stop codons, TGA and TAA. The encoded polypeptide thus includes
5 a Metionin followed by the amino acids encoded by the 136-339 fragment of the EMBL sequence J05581.

Fig. 2

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression
10 vector to give the pMRS169 expression vector. This DNA includes the sequence corresponding to nucleotides 205-720 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the
15 amino acids encoded by the 205-720 fragment of the EMBL sequence J05581.

Fig. 3

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression
20 vector to give the pMRS168 expression vector. This DNA includes the sequence corresponding to nucleotides 631-1275 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the
25 amino acids encoded by the 631-1275 fragment of the EMBL sequence J05581.

Fig. 4

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression
30 vector to give the pMRS167 expression vector. This DNA includes the sequence corresponding to nucleotides 1222-1497 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the

amino acids encoded by the 1222-1497 fragment of the EMBL sequence J05581.

Fig. 5

5 Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS175 expression vector. This DNA includes the sequence corresponding to nucleotides 136-1497 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The
10 encoded polypeptide thus includes a Metionin followed by the amino acids encoded by the 136-1497 fragment of the EMBL sequence J05581.

Fig. 6

15 Nucleotide DNA sequence (with the respective amino acid sequence) termed UBILacI. The encoded polypeptide includes the Ubiquitin sequence fused to a partial sequence of the bacterial protein beta-galactosidase, as described in Chau V. et al. *Science* 243: 1576, 1989.

Fig. 7

20 Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the expression vector pMRS30 to give the pMRS171 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence corresponding to nucleotides 136-339 of the EMBL sequence J05581
25 followed by two translation stop codons, TGA and TAA. The coded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 136-339 of the EMBL sequence J05581.

Fig. 8

30 Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS174 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 205-720 of the EMBL

sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 205-720 of the EMBL sequence J05581.

Fig. 9

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS173 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 631-1275 of the EMBL sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 631-1275 of the EMBL sequence J05581.

Fig. 10

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS172 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 1222-1497 of the EMBL sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 1222-1497 of the EMBL sequence J05581.

Fig. 11

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS176 expression vector. This DNA includes the sequence named UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 136-1497 of the EMBL sequence J05581 followed by two translation stop codons, TGA and

TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 136-1497 of the EMBL sequence J05581.

5 **Fig. 12**

Electrophoretic analysis on 1% agarose gel in 1X TBE. mRNA extracted from CHO, CD34+ dendritic cells and dendritic cells from PBMC, respectively, transfected with pMRS169, and subjected to RT-PCR reaction either with (lanes 4, 8, 12) or without (lanes 5, 9, 13) Reverse Transcriptase. Molecular weight DNA marker (lane 1); internal negative controls (lanes 2, 6); internal positive controls (lanes 3, 7, 10, 11); positive control from Promega kit (lane 14).

15 **Fig. 13**

Nucleotide sequence of the pMRS30 expression vector. The 1-2862 region corresponds to the AccI (location 504) - BamHI (location 3369) region of the pSV2CAT vector (EMBL M77788); the 2863-3721 region includes the human cytomegalovirus promoter (human cytomegalovirus major immediate-early gene enhancer); the 3722-4905 region includes several cloning sites, including XbaI (location 3727), and the processing signal of the rabbit beta-globin gene.

20 **Detailed description of the invention**

A DNA plasmid pool encoding, in eukaryotic cells, fragments of the MUC-1 human protein antigen was prepared. Constructs are based on the mammalian expression vector termed pMRS30, described in figure 13 and previously claimed in the Patent Application WO95/11982, and contain partial sequences of the MUC-1 cDNAs reported in the EMBL database with accession number J05581. MUC-1 encoding DNA was fragmented so that each fragment represents a discrete portion, partially overlapping to the adjacent ones. Administration of a mix of such plasmids can cause different plasmids to transfect different APC cells at the administration site. Therefore such cells produce and process

discrete portions of the MUC-1 protein giving the related peptides. In those conditions, the occurring subdominant and cryptic peptides can also be presented in association with class I MHC molecules thus generating a cytotoxic immune response.

5 The present invention thus relates to the use of a group of four constructs (Figures 1 to 4) containing MUC-1 cDNA partial fragments in admixture containing at least two of them and a group of four constructs (Figures 7 to 10) containing MUC-1 cDNA partial fragment preceded by the DNA encoding a protein sequence
10 containing Ubiquitin and an Escherichia coli Lac I portion (Figure 6) used separately or in admixture containing at least two of them.

 The present invention relates also to the use of the construct (Figure 5) containing the almost complete sequence of
15 the MUC-1 cDNA and the construct (Figure 11) containing the almost complete sequence of the MUC-1 cDNA preceded by the DNA encoding a protein sequence containing Ubiquitin and an Escherichia coli Lac I portion.

 The mixture of the four constructs containing the partial
20 fragments of the MUC-1 cDNA and the mixture of the four constructs containing the partial fragments of the MUC-1 cDNA preceded by the DNA encoding a protein sequence, containing Ubiquitin and an Escherichia coli Lac I portion, represents a preferred embodiment of the present invention.

25 Constructs according to the present invention can be used in the anti-tumor therapy of patient affected with tumors characterized by high MUC-1 expression.

 Constructs described in the present invention were obtained as follows.

30 In the case of the first series of constructs, the fragments of the MUC-1 DNA were obtained by RT-PCR from BT20 cell line or by DNA partial chemical synthesis. Such fragments were then cloned into the pMRS30 expression vector and verified by sequencing.

In the case of the second series of constructs, the fragments were obtained from the first series of constructs by a PCR re-amplification. These fragments were then fused to the DNA encoding the Ubiquitin (obtained by RT-PCR from MCF7 cell line mRNA) and a partial lacI sequence (obtained by PCR from the commercial vector pGEX). DNA sequences thus obtained were then cloned in the pMRS30 expression vector and verified by sequencing. For the intended therapeutic or prophylactic uses, fragments or constructs according to the invention are suitably formulated, using carriers and methods previously employed in naked DNA vaccines, as described for example in The Immunologist, 1994, 2:1; WO 90/11092, Proc. Natl. Acad. Sci. U.S.A., 1986, 83, 9551; US 5580859; Immunology today 19 (1998), 89-97; Proc. Natl. Acad. Sci. U.S.A. 90 (1993), 11478-11482; Nat. Med. 3 (1997), 526-532; Vaccine 12 (1994), 1495-1498; DNA Cell. Biol. 12 (1993), 777-783. The dosages will be determined on the basis of clinical and pharmacological-toxicological trials. Generally speaking, they will be comprised between 0.005 µg/kg and 5 µg/kg of the fragment mix. The composition of the invention can also contain a cytokine or a cytokine encoding plasmid.

The invention will be further illustrated by means of the following examples.

Example 1. Plasmid pMRS166 construction.

BT20 tumor cells (ATCC HTB-19) were cultured in Eagles MEM supplemented with 10% fetal calf serum. Ten million cells were trypsinized, washed with PBS, and mRNA extracted.

An aliquot of this RNA was subjected to RT-PCR (reverse transcriptase-polymerase chain reaction) reaction in the presence of the following synthetic oligonucleotides:

V11 (5 GATCTCTAGAATGACAGGTTCTGGTCATGCAAGC 3)

V4 (5 GATCTCTAGAAAGCTTATCAACCTGAAGCTGGTCCGTGGC 3)

The produced DNA fragment, purified and digested with the restriction enzyme XbaI, was cloned into the pMRS30 expression

vector, containing the human cytomegalovirus promoter and the beta-globin polyadenylation signal as claimed in the Patent WO9511982. The resulting pMRS166 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the nucleotides 136-339 of the EMBL sequence J05581, and two stop codons, TGA and TAA.

This fragment is reported in fig. 1.

Example 2. Plasmid pMRS169 construction.

An aliquot of the RNA obtained as reported in example 1 was amplified by RT-PCR in the presence of the following synthetic oligonucleotides:

V12 (5 GATCTCTAGAATGGTGCCAGCTCTACTGAGAAGAATGC 3)

V15 (5 GGC GGTTGGAGCCCGGGGCTGGCTTGT 3)

The produced DNA fragment, purified and digested with the restriction enzymes SmaI and XbaI, was fused, by the SmaI restriction site, to a DNA fragment entirely synthetically constructed, and including a sequence partially corresponding to the nucleotides 457-720 of the EMBL sequence J05581 and two stop codons, TGA and TAA. The whole fragment was thus cloned in the XbaI site of the pMRS30 expression vector. The resulting pMRS169 vector contains a DNA fragment including the ATG codon, the sequence partially corresponding to the nucleotides 205-720 of the EMBL sequence J05581, and two stop codons, TGA and TAA.

This fragment is reported in fig. 2.

Example 3. Plasmid pMRS168 construction.

An aliquot of the RNA obtained as reported in example 1 was amplified by RT-PCR in the presence of the following synthetic oligonucleotides:

V13 (5 GATCTCTAGAATGGGCTCAGCTTCTACTCTGGTGACAAACGGC 3)

V8 (5 GATCTCTAGAAAGCTTATCACAGGCAATGAGATAGACAATGGCC 3)

The produced DNA fragment, purified and digested with the restriction enzyme XbaI was cloned in the pMRS30 expression vector. The resulting pMRS168 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the

nucleotides 631-1275 of the EMBL sequence J05581, and two stop codons, TGA and TAA.

This fragment is reported in fig. 3.

Example 4. Plasmid pMRS167 construction.

- 5 An aliquot of the RNA obtained as reported in example 1 was subjected to RT-PCR reaction in the presence of the following synthetic oligonucleotides:

V14 (5 GATCTCTAGAATGCTGGTCTGIGTICTGGTTCGCG 3)

V10 (5 GATCTCTAGAAAGCTTATCACAAGITGGCAGAAGTGGCTGC 3)

- 10 The produced DNA fragment, purified and digested with the restriction enzyme XbaI was cloned in the pMRS30 expression vector. The resulting pMRS167 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the nucleotides 1222-1497 of the EMBL sequence J05581, and two stop
15 codons, TGA and TAA.

This fragment is reported in fig. 4.

Example 5. Plasmid pMRS175 construction.

pMRS166, 169, 168, 167 plasmids were subjected to PCR reaction in the presence of the following nucleotide pairs:

- 20 V11 (see example 1)
V18 (5 AACCTGAAGCTGGTTCGGTGGC 3) for pMRS166
V19 (5 GTGCCCAGCTCTACTGAGAAGAATGC 3)
V20 (5 GCTGGGAATTGAGAATGGAGTGCTCTTGC 3) for pMRS169
V21 (5 GGCTCAGCTTCTACTCTGGTGACACACGGC 3)
25 V22 (5 CAAGGCAATGAGATAGACAATGGCC 3) for pMRS168
V23 (5 CTGGTGCTGGTCTGTGTTCTGGTTGCG 3)
V10 (see example 4) for pMRS167

- The four DNA fragments obtained in the respective PCR reactions were mixed in equimolar amounts and PCR reacted in the
30 presence of the V11 and V10 oligonucleotides.

The produced DNA fragment, purified and digested with the XbaI restriction enzyme, was cloned in the pMRS30 expression vector. The resulting pMRS175 vector contains a DNA fragment including the ATG codon, the sequence partially corresponding to

the nucleotides 136-1497 of the EMBL sequence J05581 and two stop codons TGA and TAA.

This fragment is reported in fig. 5.

Example 6. Plasmid pMRS171 construction.

- 5 MCF7 tumor cells (ATCC HTB-22) were cultured in Eagles MEM supplemented with 10% fetal calf serum. Ten million cells were trypsinized, washed with PBS, and mRNA extracted.

An aliquot of this RNA was subjected to RT-PCR in the presence of the following synthetic oligonucleotides:

- 10 UBIup (5GATCTCTAGAATGCAGATCTTCGTGAAGACCCCTGACTGGT 3)
UBIdown
(5TCACCAGCGAGACGGGCAACAGCCATGCACCACTACCGTGCCTCCACCTCTGAGACGGAGC
ACCAAG 3)

The reaction produces a DNA fragment termed fragment 1.

- 15 DNA from pGEX11T (Pharmacia) was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

LacIup (5CCTCGTCTCAGAGGTGGGAGGCACGGTAGTGGTGCACTGGCTGTGGCC
GTCTCGCTGGTGAAAAG 3)

LacIdown (5GATCGGATCCTCGGAAACCTGTCGTGCCAGCTGC 3)

- 20 This reaction gives a DNA fragment termed fragment 2.

The 1 and 2 DNA fragments, obtained in the respective PCR reactions, were mixed in equimolar amounts and subjected to PCR reaction in presence of the UBIup and LacIdown oligonucleotides.

- 25 The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was cloned into the pUC18 commercial plasmid. The resulting pMRS156 vector contains a DNA fragment including the sequence encoding the ubiquitin fused to the sequence encoding a bacterial beta-galactosidase portion. This fragment, termed UBILacI, is reported in fig. 6.

- 30 Plasmid pMRS166 DNA was subjected to a PCR reaction in presence of the following synthetic oligonucleotides:

V3 (5GATCGGATCCACAGGTTCTGGTCATGCAAGC 3)

V4 (see Example 1)

The produced DNA fragment, purified and digested with the

restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS171 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 136-339 nucleotides of the EMBL sequence J05581 and two stop codons, TGA and TAA. This fragment is reported in fig. 7.

Example 7. Plasmid pMRS174 construction.

Plasmid pMRS169 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

V5 (5GATCGGATCCGTGCCAGCTCTACTGAGAAGATGC 3)

V6 (5GATCTCTAGAAAGCTTATCAGCTGGGAATTGAGAATGGAGTGCTCTTGC 3)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS174 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 205-720 nucleotides of the EMBL sequence J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 8.

Example 8. Plasmid pMRS173 construction.

Plasmid pMRS168 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

V7 (5GATCGGATCCGGCTCAGCTTCTACTCTGGTGCAACAGGC 3)

V8 (see example 3)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS173 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 631-1275 nucleotides of the EMBL sequence

J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 9.

Example 9. Plasmid pMRS172 construction.

5 Plasmid pMRS167 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

V9 (5 GATCGGATCCCTGGTGTCTGTCTCTGGTTCGCG 3)

V10 (see example 4)

10 The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS172 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 1222-1497 nucleotides of the EMBL sequence
15 J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 10.

Example 10. Plasmid pMRS176 construction.

20 Plasmid pMRS167 DNA was subjected PCR reaction in the presence of the following synthetic oligonucleotides:

V3 (see example 6)

V10 (see example 4)

25 The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS176 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 136-1497 nucleotides of the EMBL sequence J05581, and two stop codons, TGA and TAA. This fragment is
30 reported in fig. 11.

Example 11. Eukaryotic cell transfection and testing for transcription.

CHO (Chinese Hamster Ovary) cells were cultured in alpha MEM supplemented with ribonucleotides and deoxyribonucleotides

at transfection time.

Dendritic cells were obtained from CD34+ hemopoietic precursors cultured in IMDM without serum, supplemented with GM-CSF, IL4, SCF, Flt3 and TNFalpha. After 7 days the obtained cell population was transfected.

Dendritic cells were obtained from monocytes isolated from PBMC (peripheral blood mononuclear cells), cultured in RPMI supplemented with FCS, GM-CSF, and IL-4. After 7 days the obtained cell population was transfected.

In each case, about one million cells were transfected with one of the plasmids reported in examples 1 to 10. Transfection was carried out using 3 µg of plasmid DNA and 4 µl of DMRIE (Gibco) by lipofection.

After 24 hours cells were harvested, washed with PBS and lysed in order to extract the mRNA.

A mRNA aliquot was subjected to RT-PCR reaction in the presence of the oligonucleotide pair specific for the transfected DNA plasmid.

This experiment was carried out for each plasmid reported in the examples 1 to 10, using the following oligonucleotide pairs: V11/V4 for pMRS166, V12/V6 for pMRS169, V13/V8 for pMRS168, V4/V10 for pMRS167, V4/V10 for pMRS175, UBIup/V4 for pMRS171, UBIup/V6 for pMRS174, UBIup/V8 for pMRS173, UBIup/V10 for pMRS172, V14/V10 for pMRS176.

As a representative example, figure 12 reports the electrophoretic analysis of the DNA fragments obtained by RT-PCR from the mRNA of the three cell populations, transfected with the pMRS169 plasmid. In this case the oligonucleotide pair V12/V6 was used.

Example 12. In vivo study results.

In the *in vivo* studies, the mixtures of the four fragments and the pMRS30 plasmid (vector without insert and thus used as a negative control) were used. In order to test the occurred immunization, an ELISA test was used to show the human mucin

specific antigens.

The *in vivo* studies were conducted using human MUC1 transgenic C57BL mice. As a consequence in these animals the MUC1 protein represents a self-protein. The employed vaccination schedule consists of 3 intradermic (dorsal portion, 50 micrograms DNA for each side) administrations (at days 0, 14, 28) of 100 micrograms plasmid DNA. At day 14 after the last administration, the animals were sacrificed and sera were tested for anti-human mucin antibodies.

The assayed fragment mixes, object of the present invention, stimulated a good immune response in the treated animals.

On the other hand, vaccination experiments with a 60-aminoacid peptide corresponding to the 20 aminoacids reported in fig. 2, from location 86 to location 105, repeated three times (this peptide is termed 3XTR), were also carried out.

The two vaccinations differ in the type of the elicited antibody response. The antibody titer results much more higher in the vaccination with 3XTR. Furthermore the noticed IgG subtypes are in favor of an essentially humoral (antibody) response in the case of vaccination with 3XTR, and of a cellular response (cytotoxic) in the case of vaccination with DNA. For anti-tumor therapy, a principally cytotoxic immune response is preferable. Because the experiments were carried out on transgenic mice, in whom the human mucin is "self", we can foresee a similar response in humans. This response could justify the use, as DNA vaccines, of the compounds of the present invention in the treatment of MUC1 overexpressing human tumors.

CLAIMS

1. Pharmaceutical composition containing one or more DNA molecules, encoding fragments of a protein overexpressed in
5 tumor cells in order to induce an antitumor Ag-specific immune response, in combination with suitable excipients and adjuvants.
2. Pharmaceutical composition according to claim 1 wherein the overexpressed protein is MUC-1.
3. Pharmaceutical composition according to claim 1 or 2
10 containing at least two DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
4. Composition according to claim 3 containing at least three DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
- 15 5. Composition according to claim 4 containing at least four DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
6. Composition according to claims 3, 4 or 5 wherein the DNA sequences comprise about 200 to about 700 nucleotides, each
20 sequence being contiguous and possibly partially overlapping, from about 50 to about 150 nucleotides at the 3' and/or 5' end, to the adjacent one.
7. Pharmaceutical composition according to any claim from 2 to 6 wherein the used mixture consists of, at least, two plasmid DNA
25 molecules, each containing a DNA fragment selected from those whose sequences are described in figures 1, 2, 3, and 4.
8. Pharmaceutical composition according to claim 7 wherein the used mixture consists of the pool of plasmid DNA molecules, where each molecule contains a DNA fragment selected from those
30 whose sequences are described in figures 1, 2, 3, and 4.
9. Pharmaceutical composition according to claim 1 or 2 wherein a plasmid DNA molecule containing the sequence described in figure 5 is used.
10. Pharmaceutical composition according to claims 7, 8, or 9

wherein the used plasmid DNA molecules derive from the fusion of the pMRS30 expression vector in Fig. 13 to each sequence described in figures 1, 2, 3, 4, 5.

11. Pharmaceutical composition according to claims 2 to 6 wherein the used sequences, corresponding to single fragments of the protein, are preceded in the 5' termini by the sequence described in Fig. 6 encoding the ubiquitin and a LacI portion from Escherichia Coli.
12. Pharmaceutical composition according to claim 11 wherein the mixture consists of one or more sequences deriving from joining the pMRS30 expression vector, described in Fig. 13, to a DNA sequence selected from those described in figures 7, 8, 9, and 10.
13. Pharmaceutical composition according to claim 11 wherein the mixture consists of the totality of the sequences deriving from joining the pMRS30 expression vector to a DNA sequence selected from those described in figures 7, 8, 9, and 10.
14. Pharmaceutical composition according to claim 11 wherein the mixture consists of a sequence deriving from joining the pMRS30 expression vector to the sequence described in figure 11.
15. Pharmaceutical composition according to any preceding claims, further containing a cytokine or a cytokine encoding plasmid.
16. A plasmid DNA molecule consisting of the pMRS30 expression vector joined to a DNA sequence, encoding a MUC-1 protein fragment and whose sequence is selected from the group of those described in figures 1, 2, 3, 4, and 5.
17. A DNA molecule encoding a protein MUC-1 fragment preceded in its 5' terminus by the sequence described in Fig. 6.
18. A DNA molecule according to claim 17 selected from those described in figures 7, 8, 9, 10, and 11.
19. A plasmid DNA molecule obtained by joining the pMRS expression vector to a DNA molecule selected from those of claim 17 or 18.

20. Use of DNA molecules of claims 16-19 in the preparation of a composition with anti-tumor effect.

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Figure 1

1 ATGACAGGTTCTGGTCATGCAAGCTCTACCCAGGTGGAGAAAAG
1▶Met Thr Gl ySer Gl yHi sAl aSer Ser Thr ProGl yGl yGl uLys
46 GAGACTTCGGCTACCCAGAGAAGTTCAGTGCCAGCTCTACTGAG
16▶Gl uThr Ser Al aThr Gl nArgSer Ser Val ProSer Ser Thr Gl u
91 AAGAATGCTGTGAGTATGACCAGCGTACTCTCCAGCCACAGC
31▶LysAsnAl aVal Ser Met Thr Ser Ser Val LeuSer Ser Hi sSer
136 CCCGGTTCAGGCTCCTCCACCACTCAGGGACAGGATGTCAC'TCTG
46▶ProGl ySer Gl ySer Ser Thr Thr Gl nGl yGl nAspVal Thr Leu
181 GCCCCGCCACGGAACCAGCTTCAGGTTGATAA
61▶Al aProAl aThr Gl uProAl aSer Gl y.....

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Figure 2

1 ATGGTGCCCAGCTCTACTGAGAAGAATGCTGTGAGTATGACCAGC
1▶MetValProSerSerThrGluLysAsnAlaValSerMetThrSer
46 AGCGTACTCTCCAGCCACAGCCCCGGTTCAGGCTCCTCCACCACT
16▶SerValLeuSerSerHisSerProGlySerGlySerSerThrThr
91 CAGGGACAGGATGTCACTCTGGCCCCGGCCACGGAACCAGCTTCA
31▶GlnGlyGlnAspValThrLeuAlaProAlaThrGluProAlaSer
136 GGTTCAGCTGCCACCTGGGGACAGGATGTCACTCGGTCCCAGTC
46▶GlySerAlaAlaThrTrpGlyGlnAspValThrSerValProVal
181 ACCAGGCCAGCCCTGGGCTCCACCACCCGCCAGCCACGATGTC
61▶ThrArgProAlaLeuGlySerThrThrProProAlaHisAspVal
226 ACCTCAGCCCCGGACAACAAGCCAGCCCCGGAAGTACTGCTCCA
76▶ThrSerAlaProAspAsnLysProAlaProGlySerThrAlaPro
271 CCAGCACACGGTGTTACCTCGGCTCCGGATACCAGGCCGGCCCCA
91▶ProAlaHisGlyValThrSerAlaProAspThrArgProAlaPro
316 GGTAGTACCGCCCCCTCTGCCCATGGTGTCACTCTGCCCCGGAC
106▶GlySerThrAlaProProAlaHisGlyValThrSerAlaProAsp
361 AACAGGCCTGCATTGGGTAGTACAGCACCGCCAGTACACAACGTT
121▶AsnArgProAlaLeuGlySerThrAlaProProValHisAsnVal
406 ACTAGTGCCTCAGGCTCTGCTAGCGGCTCAGCTTCTACTCTGGTG
136▶ThrSerAlaSerGlySerAlaSerGlySerAlaSerThrLeuVal
451 CACAACGGCACCTCTGCGCGCGGACCACAACCCAGCGAGCAAG
151▶HisAsnGlyThrSerAlaArgAlaThrThrThrProAlaSerLys
496 AGCACTCCATTCTCAATTCACAGCTGATAA
166▶SerThrProPheSerIleProSer.....

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Figure 3

1 ATGGGCTCAGCTTCTACTCTGGTGCACAACGGCACCTCTGCCAGG
1▶ Met Gly Ser Ala Ser Thr Leu Val His Asn Gly Thr Ser Ala Arg
46 GCTACCACACCCAGCCAGCAAGAGCACTCCATTCTCAATTCCC
16▶ Ala Thr Thr Thr Pro Ala Ser Lys Ser Thr Pro Phe Ser Ile Pro
91 AGCCACCACTCTGATACTCCTACCACCCCTGGCCAGCCATAGCACC
31▶ Ser His His Ser Asp Thr Pro Thr Thr Leu Ala Ser His Ser Thr
136 AAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCTCCTCTC
46▶ Lys Thr Asp Ala Ser Ser Thr His His Ser Thr Val Pro Pro Leu
181 ACCTCCTCCAATCACAGCACTTCTCCCCAGTTGTCTACTGGGGTC
61▶ Thr Ser Ser Asn His Ser Thr Ser Pro Gl n Leu Ser Thr Gly Val
226 TCTTTCTTTTCTCTGTCTTTTCACATTTCAAACCTCCAGTTTAAT
76▶ Ser Phe Phe Phe Leu Ser Phe His Ile Ser Asn Leu Gl n Phe Asn
271 TCCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAGCTGCAG
91▶ Ser Ser Leu Gl u Asp Pro Ser Thr Asp Tyr Tyr Gl n Gl u Leu Gl n
316 AGAGACATTCTGAAATGTTTTGTCAGATTATAAACAAGGGGGT
106▶ Arg Asp Ile Ser Gl u Met Phe Leu Gl n Ile Tyr Lys Gl n Gly Gly
361 TTTCTGGGCTCTCCAATATTAAGTTTCAGGCCAGGATCTGTGGTG
121▶ Phe Leu Gly Leu Ser Asn Ile Lys Phe Arg Pro Gl y Ser Val Val
406 GTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAATGTCCAC
136▶ Val Gl n Leu Thr Leu Ala Phe Arg Gl u Gly Thr Ile Asn Val His
451 GACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCAGCCTCT
151▶ Asp Val Gl u Thr Gl n Phe Asn Gl n Tyr Lys Thr Gl u Ala Ala Ser
496 CGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGATGTGCCA
166▶ Arg Tyr Asn Leu Thr Ile Ser Asp Val Ser Val Ser Asp Val Pro
541 TTTCCTTTCTCTGCCAGTCTGGGGCTGGGGTGCCAGGCTGGGGC
181▶ Phe Pro Phe Ser Ala Gl n Ser Gly Ala Gly Val Pro Gl y Trp Gl y
586 ATCGCGCTGCTGGTGCTGGTCTGTGTTCTGGTTCGCTGGCCATT
196▶ Ile Ala Leu Leu Val Leu Val Cys Val Leu Val Ala Leu Ala Ile
631 GTCTATCTCATTGCCCTTGTGATAA
211▶ Val Tyr Leu Ile Ala Leu.....

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Figure 4

1 ATGCTGGTGTGGTCTGTGTTCTGTTGCGCTGGCCATTGTCTAT
1▶MetLeuValLeuValCysValLeuValAlaLeuAlaIleValTyr
46 CTCATTGCCTTGGCTGTCTGTCTCAGTGCCGCGAAGAACTACGGG
16▶LeuIleAlaLeuAlaValCysGlnCysArgArgLysAsnTyrGly
91 CAGCTGGACATCTTTCCAGCCCGGGATACCTACCATCCTATGAGC
31▶GlnLeuAspIlePheProAlaArgAspThrTyrHisProMetSer
136 GAGTACCCACCTACCACACCCATGGGCGCTATGTGCCCCCTAGC
46▶GluTyrProThrTyrHisThrHisGlyArgTyrValProProSer
181 AGTACCGATCGTAGCCCCCTATGAGAAGGTTTCTGCAGGTAATGGT
61▶SerThrAspArgSerProTyrGluLysValSerAlaGlyAsnGly
226 GGCAGCAGCCTCTCTTACACAAACCCAGCAGTGGCAGCCACTTCT
76▶GlySerSerLeuSerTyrThrAsnProAlaValAlaAlaThrSer
271 GCCAACTTGTGATAA
91▶AlaAsnLeu.....

Figure 5

1 ATGACAGGTTCTGGTCATGCAAGCTCTACCCAGGTGGAGAAAAG
 1▶ Met Thr Gly Ser Gly His Ala Ser Ser Pro Gly Gly Gly Lys
 46 GAGACTTCGGCTACCCAGAGAAGTTCACTGCCCCAGCTCTACTGAG
 16▶ Glu Thr Ser Ala Thr Glu Arg Ser Ser Val Pro Ser Ser Thr Glu
 91 AAGAATGCTGTGAGTATGACCAGCAGCGTACTCTCCAGCCACAGC
 31▶ Lys Asn Ala Val Ser Met Thr Ser Ser Val Leu Ser Ser His Ser
 136 CCCGGTTCAGGCTCTCTCCACCACTCAGGGACAGGATGTCACTCTG
 46▶ Pro Gly Ser Gly Ser Ser Thr Thr Glu Gly Glu Asp Val Thr Leu
 181 GCCCGGCCACCGAACCCAGCTTCAGGTTCACTGCGCCACCTGGGGA
 61▶ Ala Pro Ala Thr Glu Pro Ala Ser Gly Ser Ala Ala Thr Trp Gly
 226 CAGGATGTCACTCTCGTCCCAGTCACCAAGGCCAGCCCTGGGCTCC
 76▶ Glu Asp Val Thr Ser Val Pro Val Thr Arg Pro Ala Leu Gly Ser
 271 ACCACCCCGCCAGCCACGATGTCACTCAGCCCCGGACAACAAG
 91▶ Thr Thr Pro Pro Ala His Asp Val Thr Ser Ala Pro Asp Asn Lys
 316 CCAGCCCCGGGAAGTACCGCTCCACCAGCACAAGTGTACCTCG
 106▶ Pro Ala Pro Gly Ser Thr Ala Pro Pro Ala His Gly Val Thr Ser
 361 GCTCCGGATACCAAGGCCCGCCAGGTAGTACCGCCCTCTCTGCC
 121▶ Ala Pro Asp Thr Arg Pro Ala Pro Gly Ser Thr Ala Pro Pro Ala
 406 CATGGTGTACATCTGCCCGGACAACAGGCCTGCATTGGGTAGT
 136▶ His Gly Val Thr Ser Ala Pro Asp Asn Arg Pro Ala Leu Gly Ser
 451 ACAGCACCGCCAGTACACAACGTTACTAGTGCCTCAGGCTCTGCT
 151▶ Thr Ala Pro Pro Val His Asn Val Thr Ser Ala Ser Gly Ser Ala
 496 AGCGGCTCAGCTTCTACTCTGTGTCACAACGGCACCTCTGCGCGC
 166▶ Ser Gly Ser Ala Ser Thr Leu Val His Asn Gly Thr Ser Ala Arg
 541 GCGACCACAACCCAGCGAGCAAGAGCACTCCATTCTCAATTCCC
 181▶ Ala Thr Thr Thr Pro Ala Ser Lys Ser Thr Pro Phe Ser Ile Pro
 586 AGCCACCACTCTGATACTCTACCACCTTGCCAGCCATAGCACC
 196▶ Ser His His Ser Asp Thr Pro Thr Thr Leu Ala Ser His Ser Thr
 631 AAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCTCTCTCTC
 211▶ Lys Thr Asp Ala Ser Ser Thr His His Ser Thr Val Pro Pro Leu
 676 ACCTCTCCAATCACAGCACTTCTCCCCAGTTGTCTACTGGGGTC
 226▶ Thr Ser Ser Asn His Ser Thr Ser Pro Glu Leu Ser Thr Gly Val
 721 TCTTTCTTTTCTCTGTCTTTTACATTTCAAACCTCCAGTTTAAT
 241▶ Ser Phe Phe Phe Leu Ser Phe His Ile Ser Asn Leu Glu Phe Asn
 766 TCCTCTCTGGAAGATCCAGCACCGACTACTACCAAGAGCTGCAG
 256▶ Ser Ser Leu Glu Asp Pro Ser Thr Asp Tyr Tyr Glu Leu Glu
 811 AGAGACATTCTGAAATGTTTTTGAGATTATAAACAAGGGGGT
 271▶ Arg Asp Ile Ser Glu Met Phe Leu Glu Ile Tyr Lys Glu Gly
 856 TTCTGGGCTCTCCAATATTAAAGTTCAGGCCAGGATCTGGGTG
 286▶ Phe Leu Gly Leu Ser Asn Ile Lys Phe Arg Pro Gly Ser Val Val

(Continued)

Figure 5 (continued)

901 GTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAATGTCCAC
301▶ Val Gl nLeuThr LeuAl aPheArgGl uGlyThr l l eAsnVal Hi s
946 GACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCAGCCTCT
316▶ AspVal Gl uThr Gl nPheAsnGl nTyrLysThr Gl uAl aAl aSer
991 CGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGATGTGCCA
331▶ ArgTyrAsnLeuThr l l eSerAspVal Ser Val SerAspVal Pro
1036 TTTCTCTTCTCTGCCAGTCTGGGCTGGGGTGCCAGGCTGGGGC
346▶ PheProPheSer Al aGl nSer Gl yAl aGl yVal ProGl yTrpGl y
1081 ATCGCGCTGCTGGTGCTGGTCTGTGTTCTGGTTGCGCTGGCCATT
361▶ l l eAl aLeuLeuVal LeuVal CysVal LeuVal Al aLeuAl al l e
1126 GTCTATCTCATTTGCCCTGGCTGTCTGTCTCAGTGCCGCCGAAAGAAC
376▶ Val TyrLeu l l eAl aLeuAl aVal CysGl nCysArgArgLysAsn
1171 TACGGGCAGCTGGACATCTTTCCAGCCCGGATACCTACCATCCT
391▶ TyrGl yGl nLeuAsp l l ePheProAl aArgAspThr TyrHi sPro
1216 ATGAGCGAGTACCCACCTACCACACCCATGGGCGCTATGTGCC
406▶ Met Ser Gl uTyrProThr TyrHi sThr Hi sGl yArgTyrVal Pro
1261 CCTAGCAGTACCGATCGTAGCCCTATGAGAAGGTTTCTGCAGGT
421▶ ProSer Ser ThrAspArgSer ProTyrGl uLysVal Ser Al aGl y
1306 AATGGTGGCAGCAGCCTCTCTTACACAAACCCAGCAGTGGCAGCC
436▶ AsnGl yGl ySer Ser LeuSer TyrThrAsnProAl aVal Al aAl a
1351 ACTTCTGCCAACTGTGTGATAA
451▶ Thr Ser Al aAsnLeu•••••

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Figure 6

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC
1▶MetGlnIlePheValLysThrLeuThrGlyLysThrIleThrLeu
46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC
16▶GluValGluProSerAspThrIleGluAsnValLysAlaLysIle
91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT
31▶GlnAspLysGluGlyIleProProAspGlnGlnArgLeuIlePhe
136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC
46▶AlaGlyLysGlnLeuGluAspGlyArgThrLeuSerAspTyrAsn
181 ATCCAGAAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT
66▶IleGlnLysGluSerThrLeuHisLeuValLeuArgLeuArgGly
226 GCGAGGCAOGGTAGTGGTGCATGGCTGTTCCCGTCTCGCTGGTG
76▶GlyArgHisGlySerGlyAlaTrpLeuLeuProValSerLeuVal
271 AAAAGAAAAACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCC
91▶LysArgLysThrThrLeuAlaProAsnThrGlnThrAlaSerPro
316 CGCGCGTTGGCCGATTCAATTAATGCAGCTGGCAGCAGAGTTTCC
106▶ArgAlaLeuAlaAspSerLeuMetGlnLeuAlaArgGlnValSer
361 CGAGGATCC
121▶ArgGlySer

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Figure 7

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC
1▶ Met Gl n I l e Phe Val Lys Thr Leu Thr Gl y Lys Thr I l e Thr Leu
46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC
16▶ Gl u Val Gl u Pro Ser Asp Thr I l e Gl u Asn Val Lys Ala Lys I l e
91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT
31▶ Gl n Asp Lys Gl u Gl y I l e Pro Pro Asp Gl n Gl n Arg Leu I l e Phe
136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC
46▶ Ala Gl y Lys Gl n Leu Gl u Asp Gl y A rg Thr Leu Ser Asp Tyr Asn
181 ATCCAGAAAAGAGTCCACCCTGCACCTGGTGCTCCGCTCTCAGAGGT
61▶ I l e Gl n Lys Gl u Ser Thr Leu Hi s Leu Val L eu Arg Leu Arg Gl y
226 GGGAGGCACGGTAGTGGTGCATGGCTGTGCCCGTCTCGCTGGTG
76▶ Gl y A rg Hi s Gl y Ser Gl y Ala T rp Leu Leu Pro Val Ser Leu Val
271 AAAAGAAAAACCACCTGGCGCCAATACGCAAACCGCCTCTCCCC
91▶ Lys Arg Lys Thr Thr Leu Ala Pro Asn Thr Gl n Thr Ala Ser Pro
316 CGCGCGTTGGCCGATTTCATTAATGCAGCTGGCACCAGCAGGTTTCC
106▶ A rg Ala Leu Ala Asp Ser Leu Met Gl n Leu Ala Arg Gl n Val Ser
361 CGAGGATCCACAGGTTCTGGTCATGCAAGCTCTACCCAGGTGGA
121▶ A rg Gl y Ser Thr Gl y Ser Gl y Hi s Ala Ser Ser Thr Pro Gl y Gl y
406 GAAAGGAGACTTCGGCTACCCAGAGAAGTTTCAGTGCCAGCTCT
136▶ Gl u Lys Gl u Thr Ser Ala Thr Gl n Arg Ser Ser Val Pro Ser Ala
451 ACTGAGAAGAATGCTGTGAGTATGACCAGCAGCGTACTCTCCAGC
151▶ Thr Gl u Lys Asn Ala Val Ser Met Thr Ser Ser Val Leu Ser Ser
496 CACAGCCCCGGTTTCAGGCTCCTCCACCCTCAGGACAGGATGTC
166▶ Hi s Ser Pro Gl y Ser Gl y Ser Ser Thr Thr Gl n Gl y Gl n Asp Val
541 ACTCTGGCCCCGCCACGGAACCACTTCAGGTTGATAA
181▶ Thr Leu Ala Pro Ala Thr Gl u Pro Ala Ser Gl y •••••

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Figure 8

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC
1▶Met Gl n l l ePheVal LysThr LeuThr GlyLysThr l l eThr Leu
46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC
16▶Gl uVal Gl uProSerAspThr l l eGl uAsnVal LysAl aLysl l e
91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCACTTTT
31▶Gl nAspLysGl uGly l l eProProAspGl nGl nArgLeu l l ePhe
136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC
46▶Al aGlyLysGl nLeuGl uAspGl yA rgThr LeuSerAspTyrAsn
181 ATCCAGAAAAGAGTCCACCCTGCACCTGGTGTCTCCGTCTCAGAGGT
61▶l l eGl nLysGl uSer Thr LeuHl sLeuVal LeuArgLeuArgGly
226 GGGAGGCACGGTAGTGGTGCAATGGCTGTTGCCCGTCTCGCTGGTG
76▶GlyA rgHl sGlySer GlyAl aTrpLeuLeuProVal SerLeuVal
271 AAAAGAAAAACCACCTGGCGCCCAATACGCAAAACCGCTCTCCC
91▶LysArgLysThr Thr LeuAl aProAsnThr Gl nThr Al aSer Pro
316 CGCGCGTTGGCCGATTCAATATGCACTGGCAGACAGGTTTCC
106▶A rgAl aLeuAl aAspSer LeuMet Gl nLeuAl aArgGl nVal Ser
361 CGAGGATCCGTGCCAGCTCTACTGAGAAGAATGCTGTGAGTATG
121▶A rgGlySer Val ProSer Ser Thr Gl uLysAsnAl aVal SerMet
406 ACCAGCAGCGTACTCTCCAGCCACAGCCCCGGTTCAGGCTCCTCC
136▶Thr Ser Ser Val LeuSer Ser Hl sSer ProGlySer GlySer Ser
451 ACCACTCAGGGACAGGATGTCACTCTGGCCCCGGCCACGGAACCA
151▶Thr Thr Gl nGl yGl nAspVal Thr LeuAl aProAl aThr Gl uPro
496 GCTTCAGGTTCAGCTGCCACCTGGGGACAGGATGTCACTCGGTC
166▶Al aSer GlySer Al aAl aThr TrpGlyGl nAspVal Thr Ser Val
541 CCAGTCACCAGGCCAGCCCTGGGTCCACCACCCCGCCAGCCAC
181▶ProVal Thr ArgProAl aLeuGlySer Thr Thr ProProAl aHl s
586 GATGTCACTCAGCCCCGGACAACAAGCCAGCCCCGGGAAGTACT
196▶AspVal Thr Ser Al aProAspAsnLysProAl aProGlySer Thr
631 GCTCCACCAGCACACGGTGTACCTCGGCTCCGGATACCAGGCCG
211▶Al aProProAl aHl sGlyVal Thr Ser Al aProAspThr A rgPro
676 CCCCCAGGTAGTACCGCCCCCTCCTGCCCATGGTGTACATCTGCC
226▶Al aProGlySer Thr Al aProProAl aHl sGlyVal Thr Ser Al a
721 CCGGACAACAGGCCTGCATTGGGTAGTACAGCACCCGCCAGTACAC
241▶ProAspAsnArgProAl aLeuGlySer Thr Al aProProVal Hl s
766 AACGTTACTAGTGCCCTCAGGCTCTGCTAGCGGCTCAGCTTCTACT
256▶AsnVal Thr Ser Al aSer GlySer Al aSer GlySer Al aSer Thr
811 CTGGTGCAACAACGGCACCTCTGCGCGCGCGACCAACCCCGAGG
271▶LeuVal Hl sAsnGlyThr Ser Al aArgAl aThr Thr Thr ProAl a
856 AGCAAGAGCACTCCATTCTCAATTCCCAGCTGATAA
286▶Ser LysSer Thr ProPheSer l l eProSer.....

Figure 9

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC
 1▶ Met Gl n I l e Phe Val Lys Thr Leu Thr Gl y Lys Thr I l e Thr Leu
 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAGATC
 16▶ Gl u Val Gl u Pro Ser Asp Thr I l e Gl u Asn Val Lys Ala Lys I l e
 91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT
 31▶ Gl n Asp Lys Gl u Gl y I l e Pro Pro Asp Gl n Gl n Arg Leu I l e Phe
 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTCTGACTACAAC
 46▶ Ala Gl y Lys Gl n Leu Gl u Asp Gl y A rg Thr Leu Ser Asp Tyr Asn
 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCCTCCGTCTCAGAGGT
 61▶ I l e Gl n Lys Gl u Ser Thr Leu Hi s Leu Val I Leu Arg Leu Arg Gl y
 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCGCTCTCGCTGGTG
 76▶ Gl y A rg Hi s Gl y Ser Gl y Ala T rp Leu Leu Pro Val I Ser Leu Val
 271 AAAAGAAAAACCACCTGGCGCCCAATACGCAAAACCGCTCTCCC
 91▶ Lys Arg Lys Thr Thr Leu Ala Pro Asn Thr Gl n Thr Ala Ser Pro
 316 CGCGCGTTGGCCGATTTCATTAATGCAGCTGGCAGCAGAGGTTTCC
 106▶ Arg Ala Leu Ala Asp Ser Leu Met Gl n Leu Ala Arg Gl n Val Ser
 361 CGAGGATCCGGCTCAGCTTCTACTCTGGTGCACAACGGCACTCT
 121▶ Arg Gl y Ser Gl y Ser Ala Ser Thr Leu Val Hi s Asn Gl y Thr Ser
 406 GCCAGGGCTACCACAACCCAGCCAGCAGAGCACTCCATTCTCA
 136▶ Ala Arg Ala Thr Thr Thr Pro Ala Ser Lys Ser Thr Pro Phe Ser
 451 ATTCCCAGCCACCCTCTGATACTCTCTACCACCTTGCCAGCCAT
 151▶ I l e Pro Ser Hi s Hi s Ser Asp Thr Pro Thr Thr Leu Ala Ser Hi s
 496 AGCACCAAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCT
 166▶ Ser Thr Lys Thr Asp Ala Ser Ser Thr Hi s Hi s Ser Thr Val Pro
 541 CCTCTCACTCTCTCCAATCACAGCACTTCTCCCCAGTTGTCTACT
 181▶ Pro Leu Thr Ser Ser Asn Hi s Ser Thr Ser Pro Gl n Leu Ser Thr
 586 GGGGTCTCTTTCTTTTCTCTGTCTTTTTCATTTCAAACCTCCAG
 196▶ Gl y Val I Ser Phe Phe Phe Leu Ser Phe Hi s I l e Ser Asn Leu Gl n
 631 TTTAATTCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAG
 211▶ Phe Asn Ser Ser Leu Gl u Asp Pro Ser Thr Asp Tyr Tyr Gl n Gl u
 676 CTGCAGAGAGACATTCTGGAATGTTTTGTCAGATTTATAACAA
 226▶ Leu Gl n Arg Asp I l e Ser Gl u Met Phe Leu Gl n I l e Tyr Lys Gl n
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 811 TTCCACGCGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCA
 271▶ Val Hi s Asp Val Gl u Thr Gl n Phe Asn Gl n Tyr Lys Thr Gl u Ala
 856 GCCTCTCGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGAT
 286▶ Ala Ser Arg Tyr Asn Leu Thr I l e Ser Asp Val Ser Val Ser Asp
 901 GTGCCATTTCTCTTCTCTGCCCAGTCTGGGGCTGGGGTCCAGGC
 301▶ Val Pro Phe Pro Phe Ser Ala Gl n Ser Gl y Ala Gl y Val Pro Gl y
 946 TGGGCATCGCGCTGCTGGTGGTCTGTGTCTGTGTCTGGTGGCTG
 316▶ Trp Gl y I l e Ala Leu Leu Val Leu Val Cys Val Leu Val Ala Leu
 991 GCCATGTCTATCTCATTCGCTTGTGTATAA
 331▶ Ala I l e Val Tyr Leu I l e Ala Leu

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Figure 10

1 ATGCAGATCTTCGTGAAGACCCCTGACTGGTAAGACCATCACTCTC
16 Met Gl n l l e Phe Val Lys Thr Leu Thr Gl y Lys Thr l l e Thr Leu
46 GAAGTGGAGCCGAGTGACACCATTTGAGAATGTCAAGGCAAAGATC
16 Gl u Val Gl u Pro Ser Asp Thr l l e Gl u Asn Val Lys Al a Lys l l e
91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT
31 Gl n Asp Lys Gl u Gl y l l e Pro Pro Asp Gl n Gl n Arg Leu l l e Phe
136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC
46 Al a Gl y Lys Gl n Leu Gl u Asp Gl y A rg Thr Leu Ser Asp Tyr Asn
181 ATCCAGAAAGAGTCCACCCCTGCACCTGGTGTCTCCGTCTCAGAGGT
61 l l e Gl n Lys Gl u Ser Thr Leu Hi s Leu Val Leu Arg Leu Arg Gl y
226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTG
76 Gl y A rg Hi s Gl y Ser Gl y Al a Trp Leu Leu Pro Val Ser Leu Val
271 AAAAGAAAAACACCCCTGGCGCCCAATACGCAAACCGCCTCTCCC
91 Lys Arg Lys Thr Thr Leu Al a Pro Asn Thr Gl n Thr Al a Ser Pro
316 CGCGCGTTGGCCGATTTCATTAATGCAGCTGGCACGACAGGTTTCC
106 A rg Al a Leu Al a Asp Ser Leu Met Gl n Leu Al a Arg Gl n Val Ser
361 CGAGGATCCCTGGTGTCTGGTCTGTGTCTGGTTGCGCTGGCCATT
121 A rg Gl y Ser Leu Val Leu Val Cys Val Leu Val Al a Leu Al a l l e
406 GTCTATCTCATTCGCTTGGCTGTCTGTCTGTCAGTGCCGCCGAAGAAC
136 Val Tyr Leu l l e Al a Leu Al a Val Cys Gl n Cys Arg Arg Lys Asn
451 TACGGGCAGCTGGACATCTTCCAGCCCGGATACCTACCACCTCT
151 Tyr Gl y Gl n Leu Asp l l e Phe Pro Al a Arg Asp Thr Tyr Hi s Pro
496 ATGAGCGAGTACCCACCTACCACACCCATGGCGCTATGTGCCC
166 Met Ser Gl u Tyr Pro Thr Tyr Hi s Thr Hi s Gl y A rg Tyr Val Pro
541 CCTAGCAGTACCGATCGTAGCCCTATGAGAAGGTTTCTGCAGGT
181 Pro Ser Ser Thr Asp Arg Ser Pro Tyr Gl u Lys Val Ser Al a Gl y
586 AATGGTGGCAGCAGCTCTCTTACACAAACCCAGCAGTGGCAGCC
196 Asn Gl y Gl y Ser Ser Leu Ser Tyr Thr Asn Pro Al a Val Al a Al a
631 ACTTCTGCCAACTTGTGATAA
211 Thr Ser Al a Asn Leu • • • • •

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Figure 11

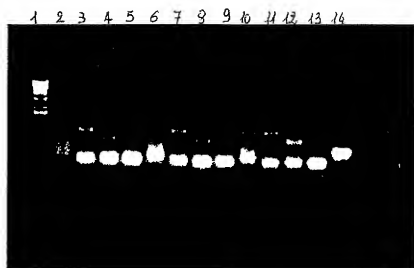
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 16▶ Gl u Val Gl u Pro Ser Asp Thr l l e Gl u Asn Val Lys Al a Lys l l e
 91 CAAGACAAGGAAGCATCCCTCCTGACCAGCAGAGGCTCATCTTT
 31▶ Gl n Asp Lys Gl u Gl y l l e Pro Pro Asp Gl n Gl n Arg Leu l l e Phe
 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC
 46▶ Al a Gl y Lys Gl n Leu Gl u Asp Gl y A rg Thr Leu Ser Asp Tyr Asn
 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT
 61▶ l l e Gl n Lys Gl u Ser Thr Leu Hi s Leu Val Leu Arg Leu Arg Gl y
 226 GGGAGGCACGGTAGTGGTGCATGGCTGTGCCCCGTCTCGCTGGTG
 76▶ Gl y A rg Hi s Gl y Ser Gl y Al a T rp Leu Leu Pro Val Ser Leu Val
 271 AAAAGAAAAACCACCTGGCGCCCAATACGCAACCCGCTCTCCC
 91▶ Lys Arg Lys Thr Thr Leu Al a Pro Asn Thr Gl n Thr Al a Ser Pro
 316 CGCGCGTTGGCCGATTTCATTATGCAGCTGGCAGCAGAGGTTTCC
 106▶ A rg Al a Leu Al a Asp Ser Leu Met Gl n Leu Al a Arg Gl n Val Ser
 361 CGAGGATCCACAGGTTCTGGTCATGCAAGCTCTACCCAGGTGGA
 121▶ A rg Gl y Ser Thr Gly Ser Gl y Hi s Al a Ser Ser Thr Pro Gl y Gl y
 406 GAAAAGGAGACTTCGGCTACCCAGAGAAGTTCAGTGCCAGCTCT
 136▶ Gl u Lys Gl u Thr Ser Al a Thr Gl n Arg Ser Ser Val Pro Ser Ser
 451 ACTGAGAAGAATGCTGTGAGTATGACCAGCAGCGTACTCTCCAGC
 151▶ Thr Gl u Lys Asn Al a Val Ser Met Thr Ser Ser Val Leu Ser Ser
 496 CACAGCCCCGGTTACGGCTCCTCCACCACCTCAGGGACAGGATGTC
 166▶ Hi s Ser Pro Gl y Ser Gl y Ser Ser Thr Thr Gl n Gl y Gl n Asp Val
 541 ACTCTGGCCCCCGGCCACGGAACACAGCTTCAGGTTTCAGCTGCCACC
 181▶ Thr Leu Al a Pro Al a Thr Gl u Pro Al a Ser Gl y Ser Al a Al a Thr
 586 TGGGGACAGGATGTACCTCGGTCCCGTCCAGTCACCAGGCCAGCCCTG
 196▶ T rp Gl y Gl n Asp Val Thr Ser Val Pro Val Thr Arg Pro Al a Leu
 631 GGCTCCACCACCCCGCCAGCCACGATGTACCTCAGCCCCGGAC
 211▶ Gl y Ser Thr Thr Pro Pro Al a Hi s Asp Val Thr Ser Al a Pro Asp
 676 AACAAAGCCAGCCCCGGGAAGTACCGTCCACCAGCACACGGTGT
 226▶ Asn Lys Pro Al a Pro Gl y Ser Thr Al a Pro Pro Al a Hi s Gl y Val
 721 ACCTCGGCTCCGGATACCAAGGCCGGCCCCAGGTAGTACCGCCCT
 241▶ Thr Ser Al a Pro Asp Thr Arg Pro Al a Pro Gl y Ser Thr Al a Pro
 766 CCTGCCCCATGGTGTACATCTGCCCCGACAAACAGGCCCTGCATTG
 256▶ Pro Al a Hi s Gl y Val Thr Ser Al a Pro Asp Asn Arg Pro Al a Leu
 811 GGTAGTACAGCACCCGCCAGTACACAACGTTACTAGTGCTCAGGC
 271▶ Gl y Ser Thr Al a Pro Pro Val Hi s Asn Val Thr Ser Al a Ser Gl y
 856 TCTGCTAGCGGCTCAGCTTCTACTCTGGTGACAAACGGCACCTCT
 286▶ Ser Al a Ser Gl y Ser Al a Ser Thr Leu Val Hi s Asn Gl y Thr Ser

(Continued)

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Figure 11 (continued)

901 GCGGCGGACCAACAACCCAGCGAGCAAGAGCACTCCATTCTCA
 301▶AlaArgAlaThrThrThrProAlaSerLysSerThrProPheSer
 946 ATTCCCAGCCACCCTCTGATACTCCTACCACCCCTTGCCAGCCAT
 316▶IleProSerHisHisSerAspThrProThrThrLeuAlaSerHis
 991 AGCACCAAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCT
 331▶SerThrLysThrAspAlaSerSerThrHisHisSerThrValPro
 1036 CCTCTCACCTCCTCCAATCACAGCACTTCTCCCCAGTTGTCTACT
 346▶ProLeuThrSerSerAsnHisSerThrSerProGlnLeuSerThr
 1081 GGGGTCTCTTTCTTTTCTCTGTCTTTTCACATTTCAAACCTCCAG
 361▶GlyValSerPhePhePheLeuSerPheHisIleSerAsnLeuGln
 1126 TTTAATTCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAG
 376▶PheAsnSerSerLeuGluAspProSerThrAspTyrTyrGlnGlu
 1171 CTGCAGAGAGACATTTCTGAAATGTTTTGCAGATTATAAACAA
 391▶LeuGlnArgAspIleSerGluMetPheLeuGlnIleTyrLysGln
 1216 GGGGGTTTTCTGGGCCCTCTCCAATATTAAGTTCAGGCCAGGATCT
 406▶GlyGlyPheLeuGlyLeuSerAsnIleLysPheArgProGlySer
 1261 GTGGTGGTACAATTGACTCTGGCCCTCCGAGAAGGTACCATCAAT
 421▶ValValValGlnLeuThrLeuAlaPheArgGluGlyThrIleAsn
 1306 GTCCACGAGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCA
 436▶ValHisAspValGluThrGlnPheAsnGlnTyrLysThrGluAla
 1351 GCCTCTCGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGAT
 451▶AlaSerArgTyrAsnLeuThrIleSerAspValSerValSerAsp
 1396 GTGCCATTTCTCTTCTCTGCCAGTCTGGGGCTGGGGTGCCAGGC
 466▶ValProPheProPheSerAlaGlnSerGlyAlaGlyValProGly
 1441 TGGGCGATCGCGCTGCTGGTCTGTGTCTGTGTCTGGTGTGCGCTG
 481▶TrpGlyIleAlaLeuLeuValLeuValCysValLeuValAlaLeu
 1486 GCCATTGTCTATCTCATTCGCTTGGCTGTCTGTCTAGTGCCCGCA
 496▶AlaIleValTyrLeuIleAlaLeuAlaValCysGlnCysArgArg
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 511▶LysAsnTyrGlyGlnLeuAspIlePheProAlaArgAspThrTyr
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 526▶HisProMetSerGluTyrProThrTyrHisThrHisGlyArgTyr
 1621 GTGCCCCCTAGCAGTACCGATCGTAGCCCCCTATGAGAAGGTTTCT
 541▶ValProProSerSerThrAspArgSerProTyrGlyLysValSer
 1666 GCAGGTAATGGTGGCAGCAGCCTCTCTTACACAAACCCAGCAGTG
 556▶AlaGlyAsnGlyGlySerSerLeuSerTyrThrAsnProAlaVal
 1711 GCAGCCACTTCTGCCAACTTGTGATAA
 571▶AlaAlaThrSerAlaAsnLeu•••••



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Figure 13

1 CCAGGAAGCTCCTCTGTGTCCTCATAAACCTAACCTCCTCTACTTGAGA
51 GGACATTCCAATCATAGGCTGCCATCCACCTCTGTGTCCTCTGTAA
101 TTAGGTCACTTAACAAAAGGAAATTGGGTAGGGGTTTTTCACAGACCGC
151 TTTCTAAGGGTAATTTTAAATATCTGGGAAGTCCCTCCACTGCTGTGT
201 TCCAGAAGTGTGTGTAACAGCCCAAAATGTCAACAGCAGAAACATACA
251 AGCTGTCAGCTTTGCACAAGGGCCCAACACCTGCTCATCAAGAAGCACT
301 GTGGTTGCTGTGTTAGTAATGTGCAAAACAGGAGGCACATTTTCCCCACC
351 TGTGTAGGTTCCAAAATATCTAGTGTTCATTCTTTACTTGGATCAGGAA
401 CCCAGCACTCCACTGGATAAGCATTTATCCTTATCCAAAACAGCCTTGTGG
451 TCAGTGTTCATCTGCTGACTGTCAACTGTAGCATTTTTTGGGGTTACAGT
501 TTGAGCAGGATATTTGGTCCTGTAGTTTGCTAACACACCTGCAGCTCCA
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601 TTTCCAGCACCATTTCATGAGTTTTTTGTGTCCCTGAATGCAAGTTTAA
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701 TCAAAATATTTCCACAGGTTAAGTCCTCATTTAAATTAGGCAAAAGGAATT
751 CTTGAAGACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTC
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851 GCGCGGAACCCCTATTGTTTATTTTCTAAATACATTCAAATATGTATC
901 CGCTCATGAGACAATAACCTGATAAATGCTTCAATAATATTGAAAAAGG
951 AAGAGTATGAGTATTCACATTTCCGTGTGCGCCCTTATTCCTTTTTTGC
1001 GGCATTTTGCCTTCCTGTTTTGTCTCACCAGAAACGCTGGTGAAAGTAA

Figure 13

2151 TAGTTAGGCCACCACTTCAAGAACTCTGTAGCACGSCCTACATACCTGCG
2201 TCTGCTAATCCTGTGTACCACTGGCTGCTGCCAGTGGCGATAAGTCGTGTG
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2951 CCATATCATAAATGTACATTTATATTGGCTCATGTCCAACATTACCGCC
3001 ATGTTGACATTGATTATTGACTAGTTATTAAATAGTAATCAATTACGGGGT
3051 CATTAGTTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTA
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3151 AATGACGTATGTTCCCATAGTAACGCCAATAGGACITTTCCATTGACGTG
3201 AATGGGTGSAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTG
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(Continued)

Figure 13 (Continued)

3301 CGCCTGGCATTATGCCAGTACATGACCTTATGGGACTTTCCTACTTGGC
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3501 GGGACTTTCAAAATGTGTAACTCCGCCCCATTGACGCAATGGGC
3551 GGTAGGCGTGTAACGTGGGAGGTCTATATAAGCAGAGCTCGTTAGTGAA
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3651 GACACCGGGACCGATCCAGCCTCCGCGGCGGGAACGGTGCAATGGAAAG
3701 CGGATTCCCGGTGCCAAGAAAGCTTGCTAGAACCCGGGAGAGCTCCTGA
3751 GAACTTCAGGGTGAGTTTGGGGACCCCTTGATTGTTCTTTCTTTTCGCTA
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4251 TGTTTGAGATGAGGATAAAATACTCTGAGTCCAAACGGGCCCTCTGCT
4301 AACCATGTTTCATGCCCTTCTCTTTTTCTACAGCTCCTGGGCAACGTGCT
4351 GGTGTTGTGCTGTCTCATCATTTTGGCAAAGAATCACTCCTCAGGTGC
4401 AGGCTGCCTATCAGAAGGTGGTGGCTGGTGTGGCCAATGCCCTGGCTCAC

(Continued)

Figure 13 (Continued)

1051 AAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAACTGGAT
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1251 GACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCTTACGGATGGCAT
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2001 TTTCTGCGCGTAATCTGCTGCTTGCAACAAAAAACCAACCGTACCAGC
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(Continued)

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Figure 13 (Continued)

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<210> 7

<211> 579

<212> DNA

<213> human

<400> 7

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 <213> human

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 <211> 1020
 <212> DNA
 <213> human

<400> 9
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<211> 651

<212> DNA

<213> human

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<210> 11

<211> 1737

<212> DNA

<213> human

<400> 11

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<211> 4905

<212> DNA

<213> human

<400> 12

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4905

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<211> 31

<212> DNA

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<223> Description of Artificial Sequence: synthetic
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31

<210> 14

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<212> DNA

<213> Artificial Sequence

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oligonucleotide

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41

<210> 15

<211> 36

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 15

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36

<210> 16

<211> 49

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
oligonucleotide

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49

<210> 17

<211> 40

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
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<210> 18

<211> 45

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
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<210> 19

<211> 38

<212> DNA

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38

<210> 20

<211> 41

<212> DNA

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41

<210> 21

<211> 34

<212> DNA

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<223> Description of Artificial Sequence: synthetic
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gatctctaga atgacaggtt ctggctcatgc aagc

34

<210> 22

<211> 39

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
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39

<210> 23

<211> 43

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
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<400> 23

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43

<210> 24
<211> 41
<212> DNA
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oligonucleotide

<400> 24
gatctctaga atgctggtgc tggctctgtgt tctggttgcg c 41

<210> 25
<211> 26
<212> DNA
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<220>
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oligonucleotide

<400> 25
ggcggtagag cccggggctg gcttgt 26

<210> 26
<211> 22
<212> DNA
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<220>
<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 26
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<210> 27
<211> 26
<212> DNA
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<223> Description of Artificial Sequence: synthetic

oligonucleotide

<400> 27

gtgccagct ctactgagaa gaatgc

26

<210> 28

<211> 29

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 28

gctgggaatt gagaatggag tgctcttgc

29

<210> 29

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 29

ggctcagctt ctactctggt gcacaacggc

30

<210> 30

<211> 25

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 30

caaggcaatg agatagacaa tggcc

25

<210> 31

<211> 27

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 31

ctggtgctgg tctgtgttct ggttgcg

27

<210> 32

<211> 40

<212> DNA

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<223> Description of Artificial Sequence: synthetic
oligonucleotide

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40

<210> 33

<211> 68

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 33

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gcaccagg 68

<210> 34

<211> 66

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
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<400> 34

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gaaaag

66

<210> 35

<211> 35

<212> DNA

<213> Artificial Sequence

<220>

<221> Description of Artificial Sequence: synthetic
oligonucleotide

<400> 35

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35